

Natural Calanolides and Their Chemical Biology and Pharmacology on Human Immunodeficiency Virus-1 and *Mycobacterium Tuberculosis*

Ran Mu¹, Tao Ma², Purong Zheng¹, Somersan-Karakaya³, Hongyao Wang¹, Xiaofan Lu⁴, Zhiwei Chen⁵, Gang Liu^{1*}

¹Department of School of Pharmaceutical Sciences, Tsinghua University, Beijing 100084, China

²Department of School of Chinese Pharmacy, Beijing University of Chinese Medicine, Beijing 100102, China

³Department of Medicine, Division of Infectious Diseases, Weill Cornell Medical College, 1300 York Avenue, New York, New York, 10065, United States

⁴Beijing Key Laboratory for HIV/AIDS Research, Center for Infectious Diseases, Beijing You'an Hospital, Capital Medical University, Beijing 100069, China

⁵AIDS Institute and Department of Microbiology, The University of Hong Kong Li Ka Shing Faculty of Medicine, Rooms 40-46, 5/F, Laboratory Block, 21 Sassoon Road, Pokfulam, Hong Kong

***Corresponding author:** Professor Gang Liu: Department of School of Pharmaceutical Sciences, Tsinghua University, Beijing 100084, China, E-mail: gangliu27@mail.tsinghua.edu.cn

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1. Description and Topical Outline

This review aims to be a comprehensive, authoritative, critical, and accessible review of general interest to the chemistry community because a natural (+)-calanolide A and its both natural and synthetic analogues were continuously investigated. (+)-calanolide A was identified in 1992 as a distinguished class of NNRTI for HIV-1 with its unique biological and pharmaceutical properties and subsequently was reported on its anti drug-susceptible and drug-resistant strains of *Mtb* in 2004. Due to its structural diversity and pharmacological potentials, readers may not only realize the importance of natural product and relative organic and medicinal chemistries for drug development, but may be also interesting in insight of the efforts on biological target identifications, druggable profile investigation (i.e. structure-activity relationship, structure-metabolism relationship and structure-toxicity relationship), pharmaceutical sciences and the translation into clinical application, i.e. as a novel diagnostic agent for TB.

2. Outline of Planned Sections

2.1. Where are calanolides from and their structure diversity?

2.2. The total synthesis of natural (+)-Calanolide A and its derivatives

2.3. Natural 1 was active against HIV-1 as NNRT inhibitor, and its clinical investigation

2.3.1. Anti-HIV-1 activity of calanolides

2.3.1.1. Anti-HIV-1 activity of calanolides in cell culture

2.3.1.2. Anti-HIV-1 activity of calanolides in hollow fiber mouse

2.3.2. Anti-HIV-1 activity of inophyllums

2.3.3. Anti-HIV-1 activity of cordatolides

2.3.4. Investigation of 1 in Clinical Trails

2.4. Improved calanolides as anti-HIV-1 NNRT inhibitor

2.5. (+)-Calanolide A and its derivatives inhibit replicating and non-replicating *M. Tuberculosis* (*Mtb*)

2.5.1. Identification of 1 as a naturally occurring anti-tubercular compound

2.5.2. Preliminary studies of structural modification based on 1

2.5.3. Identification and optimization of synthetic calanolides with nitrofuranyl moiety active against replicating and non-replicating *Mtb*.

2.5.4. Other modifications of the calanolides aimed for assay of anti-tuberculosis

2.6. Calanolide nitrofuranyl derivatives (NFCs) target at a nitro reductase Rv2466c requiring mycothiol as a cofactor

2.7. Development of an Rv2466c-dependent fluorescent probe for *Mtb* diagnosis and drug susceptibility testing

2.8. Investigation of the interaction with Rv2466c and NFCs compound in the presence of MSH

2.8.1. Reduction of NFCs by Rv2466c requires an active conformation induced by mycothiolation

2.8.2. The role of MSH

2.8.2.1. Synthesis of MSH and its derivatives

2.8.2.2. The contribution of different MSH's motifs in activation of Rv2466c

2.8.2.3. Reduction of NFCs requires an active conformation of Rv2466c via mycothiolation

2.9. NFCs developed as a novel fluorescent probe for rapid and high throughput diagnosis of TB

2.9.1. The dilemma of treatment and diagnosis of TB and current diagnostic and DST methods in hospitals and research labs

2.9.2. NFCs developed as a novel fluorescent probe to detect *Mtb*

2.9.2.1. The specificity and sensitivity of NFCs-based *Mtb* detected method

2.9.2.2. Detection of clinical sputum specimens from out-patients by the NFC method

2.9.2.3. Evaluation of pDST by the NFC method in a high-throughput manner

2.10. Summary

chloromethyl-11-demethyl-12-oxo calanolide A with druggable profile. *J. Med. Chem.*, 2010, 53: 1397-1401.

(3) Lu, X. F.; Liu, L.; Zhang, X.; Lau, T. C. K.; Tsui, S. K. W.; Kang, Y. X.; Zheng, P. R.; Zheng, B. J.; Liu, G.; Chen, Z. W. F18, a novel small-molecule NNRTI, inhibits HIV-1 replication using distinct

binding motifs as demonstrated by resistance selection and docking analysis. *Antimicrob. Agents Chemother.*, 2011, 341-351.

(4) Zheng, P. R.; Somersan-Karakaya, S.; Lu, S. C.; Roberts, J.; Pingle M.; Warriar, T.; Little, D.; Guo, X. Y.; Brickner, S. J.; Nathan, C. F.; Gold B.; Liu, G. Synthetic calanolides with bactericidal activity against replicating and non-replicating Mycobacterium tuberculosis, *J. Med. Chem.* 2014, 57, 3755-3772.

(5) Ran Mu, Chengcheng Kong, Wenjun Yu, Hongyao Wang, Jie Wu, Xueyuan Li, Selin Somersan-Karakaya, Haitao Li, Zhaogang Sun, Gang LIU, Nitrooxidoreductase Rv2466c-Dependent Fluorescent Probe for Mycobacterium tuberculosis Diagnosis and Drug Susceptibility Testing, *ACS Infect. Dis.* 2019, 5(6), 949-961.

3. List of Previous Reviews of the Subject

1. Newman DJ, Cragg GM. Natural Products as Sources of New Drugs from 1981 to 2014. *J Nat Prod.* 2016; 79(3): 629-61. doi: 10.1021/acs.jnatprod.5b01055.

2. Gu SX, Xiao T, Zhu YY, Liu GY, Chen FE. Recent progress in HIV-1 inhibitors targeting the entrance channel of HIV-1 non-nucleoside reverse transcriptase inhibitor binding pocket. *Eur J Med Chem.* 2019; 174: 277-291. doi: 10.1016/j.ejmech.2019.04.054.

3. Nguta JM, Appiah-Opong R, Nyarko AK, Yeboah-Manu D, Addo PGA. Current perspectives in drug discovery against tuberculosis from natural products. *Int. J. Mycobacteriol.* 2015; 4(3): 165-83. doi: 10.1016/j.ijmyco.2015.05.004.

4. angeby K, Juréen P, Kahlmeter G, Hoffne SE & Schön T. Challenging a dogma: antimicrobial susceptibility testing breakpoints for Mycobacterium tuberculosis. *Bull World Health Organ.* 2012; 90:693-698. doi:10.2471/BLT.11.096644.

4. List of Five of the Authors' Papers that are Most Closely Related to the Review Topic

1. Ma T, Liu L, Xue H, Li L, Han CY, Wang L, et al. Chemical library and structure-activity relationships of 11-demethyl-12-oxo calanolide A analogues as anti-HIV-1 agents. *J. Med. Chem.*, 2008, 51: 1432-1446.

2. Xue H, Lu XF, Zheng PR, Liu L, Han CY, Hu JP, et al. Highly suppressing wild-type HIV-1 and Y181C mutant HIV-1 strains by 10-chloromethyl-11-demethyl-12-oxo calanolide A with druggable profile. *J Med Chem.* 2010; 53: 1397-1401.

3. Lu XF, Liu L, Zhang X, Lau TCK, Tsui SKW, Kang YX, Zheng PR, Zheng BJ, Liu G, Chen Z. F18, a novel small-molecule NNRTI, inhibits HIV-1 replication using distinct binding motifs as demonstrated by resistance selection and docking analysis. *Antimicrob. Agents Chemother.* 2011; 56(1): 341-351.

4. Zheng PR, Somersan-Karakaya S, Lu SC, Roberts J, Pingle M, Warriar T, et al. Synthetic calanolides with bactericidal activity against replicating and non-replicating *Mycobacterium tuberculosis*. *J Med Chem.* 2014; 57: 3755-3772.

5. Ran Mu, Chengcheng Kong, Wenjun Yu, Hongyao Wang, Jie Wu, Xueyuan Li, et al. Nitrooxidoreductase Rv2466c-Dependent Fluorescent Probe for *Mycobacterium tuberculosis* Diagnosis and Drug Susceptibility Testing. *ACS Infect. Dis.* 2019; 5(6): 949-961.